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1 Natural diversity in the carotene, tocochromanol and fatty acid composition

2 of crude palm oil

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17 Highlights

- 18 Considerable intraspecific diversity exists in most crude palm oil compounds
- 19 The geographical origin of oil palm significantly influences oil composition
- Palms from Côte d'Ivoire showed the lowest palmitate and highest vitamin E levels
- High (>500 ppm) carotene contents were found in various origins in West Africa
- Hybrids of Côte d'Ivoire × Nigeria crosses produce oil of high nutritional value
- 23
- 24

25 Abstract

26 Crude palm oil (CPO) is extracted from the mesocarp of oil palm (Elaeis guineensis) fruits. CPO is 27 widely consumed in many African countries. Due to its high provitamin A carotenoid content, it is 28 also widely used in programmes designed to prevent vitamin A deficiency. Elaeis guineensis occurs 29 naturally across a wide geographical range in Africa. We investigated the carotene, tocochromanol 30 (vitamin E) and fatty acid composition of a large set of genotypes representative of this genetic and 31 geographic diversity. We found considerable intraspecific diversity in most lipid traits. Populations 32 from Côte d'Ivoire were distinguished from other origins by their very low palmitate content and high 33 tocochromanol content. Genotypes from Benin, Côte d'Ivoire and Nigeria were characterized by high 34 carotene contents. Finally, hybrids of crosses between genotypes from Côte d'Ivoire and Nigeria 35 produce CPO with exceptionally high provitamin A and vitamin E contents together with low 36 palmitate content.

37

38 Keywords

39 Vitamin E; Provitamin A; Palmitic acid; Oil palm, Red palm oil; Geographical origin

40

41 **1. Introduction**

42 Crude palm oil (CPO), also called red palm oil, is extracted from the mesocarp of *Elaeis guineensis* 43 fruits. CPO has been consumed in Africa for thousands of years (Kay & Kaplan, 2015) and is still 44 widely used in many West African countries (Rafflegeau, Nanda, & Genot, 2018), where it can 45 represent up to 60% of edible oil consumption, as is the case in Cameroon (Rébéna, Rafflegeau, 46 Kanski, Nanda, & Genot, 2019). In countries where CPO is traditionally used for everyday cooking, 47 consumers look for deep colour and high fluidity (Cheyns, Bricas, & Aka, 2004).

48 The two main pigments responsible for the colour of CPO, α - and β -carotenes, are also 49 provitamin A carotenoids. CPO is one of the richest dietary sources of provitamin A and is thus also 50 used in programmes to prevent long-term vitamin A deficiency (Zagré, Delpeuch, Traissac, & Delisle, 51 2003; Burri, 2012; Delisle, 2018). Vitamin A deficiency is a public health concern in numerous 52 developing countries. It can affect up to 80% of the population in sub-Saharan countries and 50% in 53 South East Asia (Murphy, 2007; Hamer & Keusch, 2015). Vitamin A deficiency has a plethora of 54 clinical manifestations (night blindness, growth disturbances, and susceptibility to severe infection) 55 and increases the mortality of young children and mothers (Black et al., 2008). Worldwide, more than 56 half a million children under the age of five die every year due to vitamin A deficiency. The 57 identification of oil palm varieties with a high carotene content could thus not only satisfy consumer 58 demand but also help prevent vitamin A deficiency.

59 A comprehensive literature review revealed that very little is known about intraspecific variability 60 of the carotenoid content of CPO. Most reviews (e.g., Sundram, Sambanthamurthi, & Tan, 2003) 61 provide the same figures for CPO total carotenoid content, i.e. 500-700 ppm, but a closer 62 examination showed that this range of values was based on very few studies. However, a few original 63 studies did show that variations in CPO carotenoid content between oil palm origins can be 64 significantly higher. Among four Nigerian genotypes, carotenoid content was found to vary from 648 65 to 2 226 ppm (Ames, Raymond, & Ward, 1960). Monde et al. (2009) found a higher carotenoid content (1 532 ppm) in a variety originating from Côte d'Ivoire than in a variety belonging to the Deli 66 group (690 ppm). Finally, a range of 450 to 1 250 ppm was reported among 42 progenies collected in 67 68 Angola (Noh et al., 2002).

69Because the specific vitamin activity and bioavailability of the different carotenoids varies70considerably (Haskell, 2012), carotenoid composition also warrants attention. In the few original71studies available, β-carotene and α-carotene were always the two major carotenoids in CPO. β-72carotene, which has twice the vitamin A potential of α-carotene, represented 86% of carotenoids in a73variety from Côte d'Ivoire (Monde et al., 2009) but 60% and 72% in two Nigerian varieties (Ikemefuna)

8 Adamson, 1984), suggesting the ratio of β-carotene to α-carotene may also vary among oil palm
 75 from different origins in Africa.

76 CPO is also well known for its high vitamin E content (Schroeder, Becker, & Skibsted, 2006; Teh & 77 Lau, 2021). Tocochromanols are a group of four tocopherols and four tocotrienols that collectively 78 constitute vitamin E, an essential micronutrient in the human diet. A specific feature of CPO is its 79 particularly high concentration of tocotrienols, which may account for 80-95% of total 80 tocochromanols (Monde et al., 2009; Irias-Mata et al., 2017). Tocotrienols are the subject of 81 increasing interest because of their health properties (Trela & Szymanska, 2019; Wallert, Börmel, & 82 Lorkowski, 2021). Because tocochromanols and carotenes protect each other from oxidation during 83 storage or cooking (Schroeder, Becker, & Skibsted, 2006), identifying oil palm origins with high 84 tocochromanol content is not only important as a source of vitamin E but also for provitamin A 85 intake. Again, reviews of CPO tocochromanol contents are based on a limited literature and report 86 different ranges: 500-800 ppm (Sundram, Sambanthamurthi, & Tan, 2003) or 600-1 000 ppm (Hoe, 87 Chan, Ramanan, & Ooi, 2020; Goh, Choo, & Ong, 1985). Original studies are rare but provide a 88 different picture of intraspecific variability of CPO vitamin E content. Luo et al. (2020) recently found 89 values ranging between 172 and 1 288 ppm in a set of 161 oil palm individuals collected in different 90 producer countries. Unfortunately, the African origin of the genotypes they studied was not 91 investigated. Low tocochromanol contents (332 to 525 ppm) were found in three varieties grown in 92 Costa Rica (Irias-Mata et al., 2017), but high vitamin E content (1 124 ppm) was found in the 93 genotype of the Deli group studied by Monde et al. (2009).

94 Palm oil is known for its high level of palmitic acid (16:0), meaning it has many advantages for the 95 food industry, including high oxidative stability and a high melting point, making it a good alternative 96 to trans fats (hydrogenated oils). The range of 16:0 content usually reported for palm oil is 39-46% 97 (e.g., Sundram, Sambanthamurthi, & Tan, 2003). However, CPO consumers in Africa want their CPO 98 to be fluid at ambient temperature (Cheyns, Bricas, & Aka, 2004), i.e., a CPO with a low 16:0. Little is 99 known about the intraspecific variability of fatty acid composition in E. guineensis. However, a few 100 studies reported that oil palms originating from Côte d'Ivoire may have a 16:0 level lower than 39% 101 (Gascon & Wuidart, 1975; Monde et al., 2009; Montoya et al., 2014).

Our knowledge of the variability of CPO carotene, tocochromanol and FA levels linked with the different geographic origins and genetic groups of oil palm thus remains very limited. Oil palm is a native of west and central Africa, from Senegal to Angola along the Atlantic Ocean, and as far as Tanzania in the east. Genetic analyses revealed two major genetic groups within *E. guineensis*: Group l contains collections from Senegal, Guinea, Sierra Leone, and Côte d'Ivoire, while Group II includes all other collections, including those from Benin, Nigeria, Cameroon, Congo, and Angola (Cochard et 108 al., 2009; Bakoumé et al., 2015). A third genetic group, termed Deli, which is of considerable 109 importance for oil palm breeding, contains material introduced from Africa and bred in Indonesia in 110 the middle of the 19th century, but the area of origin of this material in Africa remains unknown 111 (Cochard et al, 2009; Bakoumé et al, 2015). The aim of this study was thus to characterize carotene, 112 tocochromanol and FA composition of CPO in a large set of genotypes representing the three genetic 113 groups of oil palm. We analysed 44 genotypes from Angola, Benin, Cameroon, Congo, Côte d'Ivoire, 114 Nigeria, and the group Deli, and assessed diversity within and between origins in each of the three 115 classes of lipid compounds. Twelve inter-group hybrids were further analysed to test whether new 116 genetic combinations could provide CPO with high provitamin A and vitamin E contents and a low 117 16:0 level.

118

119 **2. Material and methods**

120 2.1. Plant material and crude oil extraction in Benin

121 All the genotypes studied were grown at the Agricultural Research Centre for Perennial Plants 122 (Centre de Recherches Agricoles Plantes Pérennes, CRA-PP/INRAB) at Pobè, Benin. Passport data 123 (genotype identifier, country and population of origin, genetic group, planting year) of each genotype 124 are given in Supplementary Table 1. The genetic group of each genotype was determined in a 125 previous study (Cochard et al., 2009). We studied 10 genotypes originating from Côte d'Ivoire 126 belonging to two populations (La Mé, seven genotypes, and Yocoboué, three genotypes), five 127 genotypes from Benin (population Pobè), four genotypes from Nigeria (population Aba-Calabar), five 128 genotypes from Congo (population Yangambi), four genotypes from Angola, belonging to two 129 populations (Cuanza and N'Jala, two genotypes each), and 16 genotypes of the group Deli, divided 130 into two sub-groups (Dabou, 13 genotypes) and (Socfin, three genotypes) (Fig. 1). The 12 inter-group 131 hybrids investigated were chosen because at least one of their two parents did not belong to any of the nine populations (including the two Deli sub-groups) in the CRA-PP field genebank (Fig. 1). For 132 133 each genotype, the objective was to extract and analyse CPO in triplicate from three independent 134 bunches, i.e., collected on three different dates in the period 2017-2019. Each genotype 135 corresponded to single palm tree in the CRA-PP field genebank. In the end, four bunches were 136 harvested and analysed in six genotypes: three bunches in 47 genotypes and only two bunches in 137 three genotypes (Supplementary Table 1). Forty well-developed undamaged fruits were removed 138 from spikelets collected in the centre of each bunch. The fruits were immediately immersed in 139 boiling water for 20 min or until a final temperature of 75-80 °C was reached in the centre of three 140 fruits per bunch measured with a Stuart SCT1 probe thermometer (Cole-Parmer, Stone, UK). After

heating, fruits were left to cool to 60 °C (internal mesocarp temperature) and then pressed using a manual 1.2 L screw-press. The extracted crude oil was stirred while still warm and fluid, and 10 mL were sampled and immediately frozen at -20 °C. At this stage of the process, crude oil still contains water and impurities. Unpurified crude oil samples were stored at -20 °C at Pobè for maximum two months.

146

147 2.2. CPO purification

148 Unpurified crude oil samples were sent to Montpellier, France, by plane and kept cold during 149 transport using eutectic plates. Prior to lipid extraction, after 15 min in a 40 °C water-bath (Firlabo, 150 Meyzieu, France) to melt all the triacylglycerols, the samples were homogenised. Total lipids were 151 extracted from the samples of crude oil using a modified Folch method as described previously 152 (Laffargue, de Kochko, & Dussert, 2007). Briefly, 3 mL of crude oil was homogenised for 30 s in 20 mL 153 of methylene chloride/methanol (2/1) (analytical grade, Merck, Darmstadt, Germany) using a IKA T25 154 Ultra Turrax homogeniser (Staufen, Germany) prior to filtration (pore size 5) into a glass filter cup. 155 The filtrate was transferred in a 100 mL glass separatory funnel and washed with 5 mL of 0.73% NaCl solution by vigorous hand shaking. After the resulting mixture separated into two phases, the lower 156 157 phase was recovered and divided into nine aliquots, i.e. three aliquots for each of the three classes of 158 compounds to be analysed (carotenoids, tocochromanols and fatty acids).

159

160 2.3. Carotenoid analysis

161 Carotenoid extraction was adapted from the method described previously (Montúfar et al., 2010), 162 which includes a preliminary triacylglycerol (TAG) removal step. Briefly, CPO samples (15 mg) dried 163 under nitrogen (≥ 99.999%, Linde, Lyon, France), using an OASys heating system (Organomation, 164 Berlin, MA, USA) were carefully mixed with 10 mL of acetone (analytical grade, Merck) and left for 60 165 min at -20 °C, leading to TAG crystallisation. After centrifugation (Eppendorf 5810R, Hamburg, 166 Germany) of frozen samples at 900 g and -9 °C for 20 min, TAG were separated by rapid sampling of 167 the upper acetone phase, which contained the carotenoids, and filtered through a 0.2 µm Minisart 168 SRP4 PTFE filter (Sartorius, Göttingen, Germany). The 20-µL carotenoid extract was directly injected 169 into the Agilent Infinity II HPLC system (Santa Clara, CA, USA). The column was a polymeric YMC-30 170 (300 × 4.6 mm, YMC, Kyoto, Japan). Elution was performed at a flow rate of 1 mL/min as follows: (i) 171 100% A for 1 min, (ii) a linear gradient from 100% A to 10% A and 90% B for 40 min, (iii) a linear 172 gradient to 100% B for 5 min, (iv) a 5-min step with 100% B, and (v) a return to initial conditions for 5 173 min (A = methanol:water [60:40, v/v], B = methanol:methyl tert-butyl ether:water [28.5:67.5:4,

174 v/v/v]). A 15-min rinse step was then carried out using 100% ethyl acetate, followed by re-175 equilibration with 100% A for 10 min. All solvents were of HPLC grade and were purchased from 176 Merck. The column temperature was maintained at 25 °C. An Agilent G7115A UV-visible photodiode 177 array detector was used to analyse the chromatograms at a detection wavelength of 450 nm. 178 Carotenoids were identified by the combined use of their relative retention times and their 179 wavelength absorption maxima determined using analytical standards (Merck). All analyses were 180 performed in triplicate from three independent carotenoid extractions.

181

182 2.3. Tocochromanol analysis

183 Analysis of tocochromanols was adapted from the method described previously (Montúfar et al., 184 2010). CPO samples (15 mg) dried under nitrogen were carefully dissolved in 500 μ L of *n*-hexane and 185 30 µL aliquots of the solution were injected into the Agilent Infinity II HPLC system, which was 186 equipped with an Agilent G7121B fluorescence detector set at an excitation wavelength of 290 nm 187 and an emission wavelength of 330 nm. The HPLC column was a Merck Lichrospher Si-60 (250 × 4.6 188 mm internal diameter, 5 μ m particle size). The mobile phase was *n*-hexane/propan-2-ol (99:1, v/v) at 189 a flow rate of 1 mL/min. All the solvents were of HPLC grade and were purchased from Merck. 190 Standard curves were run with commercial standards of α -, β -, γ - and δ -tocopherols (Tocopherol set, 191 Calbiochem, Merck) and α -, β -, γ - and δ -tocotrienols (Cayman Chemical, Ann Harbor, MI, USA). All 192 analyses were performed in triplicate from three independent CPO dissolutions in *n*-hexane.

193

194 2.4. Fatty acid analysis

195 Fatty acid methyl esters (FAME) were prepared according to the ISO-5509 standard from 50 mg CPO 196 samples and analysed using gas chromatography to determine FA composition (Laffargue, de Kochko, 197 & Dussert, 2007). Analyses were performed using an Agilent 7820A Gas Chromatography system with 198 flame ionization detection. A Famewax capillary column (Restek, Lisses, France), (30 m × 0.25 mm × 199 0.25 m) was used. Analyses were conducted from 185 to 225 °C at 4 °C/min and then kept at 225 °C 200 for 10 min. The carrier gas was helium at 40 cm/s. Both injector and detector were maintained at a 201 temperature of 230 °C. Only peaks identified in the chromatograms with a relative percentage >0.1% 202 were retained. All fatty acid analyses were performed in triplicate from three independent FAME 203 reactions.

204

205 2.5. Statistical analyses

206 Genotype means were calculated using two to four biological replicates (two to four bunches 207 collected from the same palm tree on different dates) and three technical replicates per biological 208 replicate. Linear regression and one-way ANOVA were performed using Statistica (Statsoft, Tulsa, 209 USA). Linear regression was used to test the relationships between carotene content, tocochromanol 210 content, the β -carotene to α -carotene ratio, and the percentages of tocotrienols, linoleic acid and α -211 and β -tocochromanol forms. ANOVA was used to test the effect of the country of origin and the 212 population on all chemical traits measured. When the effect of the country of origin or the 213 population was significant, Duncan's post-hoc multiple comparison test was performed using 214 Statistica. The accuracy of the classifications based on carotene, tocochromanol and fatty acid 215 compositions was also tested using discriminant analysis (DA) and canonical analysis (CA). DA and CA 216 were performed using the following active variables: total carotene content, β -carotene to α -217 carotene ratio, total tocochromanol content, the percentage of tocotrienols, VTE3 and VTE4 218 activities, and the percentages of 14:0, 16:0, 16:1, 18:0, 18:1n-7, 18:2, 18:3, 20:0 and 20:1n-9. The 219 population or the country of origin was the grouping variable. DA and CA were performed using 220 Statistica.

221

222 **3. Results and Discussion**

223 3.1. Intraspecific variability of carotene, tocochromanol and fatty acid composition of crude palm oil

224 Total carotene content varied considerably in the 44 genotypes chosen to represent the geographic 225 and genetic diversity of oil palm: from 77 ppm in the genotype Yang5 to 1 275 ppm in the genotype 226 Pobe3 (Fig. 2a; Supplementary Table 2). The CPO carotene content of 16 out of 44 genotypes was 227 above the limit of the range usually reported in the literature (500-700 ppm), thus supporting the 228 possible identification of high provitamin A genotypes, as previously reported by Monde et al. (2009) 229 and Noh et al. (2002). More unexpectedly, we found that more than half the genotypes studied 230 contained less than 500 ppm of carotenes. In five genotypes, CPO carotene content was even less 231 than 200 ppm. We therefore advise caution concerning the origin of the CPO when calculating 232 recommended CPO intakes for use in programmes to prevent vitamin A deficiency (Burri, 2012).

233 In all the genotypes studied here, α -carotene and β -carotene were the only two major 234 carotenoids. None of the other minor carotenoids (< 0.5%) reported in the literature (Sundram, 235 Sambanthamurthi, & Tan, 2003) was identified as a major compound in our large sample set, 236 suggesting that the intraspecific diversity in the different types of carotenoids accumulated in the 237 mesocarp of oil palm fruits is very restricted. In contrast, although the majority of genotypes had the 238 β -carotene to α -carotene ratio usually given for CPO, i.e., between 1.5 and 2, the ratio varied 239 considerably among the 44 genotypes studied (Fig. 2b). In the genotype Pobe5, β -carotene content 240 was 7 times higher than that of α -carotene (518 vs 75 ppm, Supplementary Table 2). Such a high ratio 241 was also observed in one of the two origins studied by Monde et al. (2009). Conversely, in the 242 genotype AbaC1, α -carotene represented 60% of the CPO carotenoids. Considering that the vitamin 243 A equivalence of α -carotene is half that of β -carotene (Haskell, 2012), the choice of the source of 244 CPO also appears to be crucial regarding these characteristics. Finally, it is worth noting that the β -245 carotene to α -carotene ratio was not correlated with the total carotene content of the CPO (Fig. 3).

246 Total tocochromanol content also varied considerably within the panel of genotypes investigated 247 (Fig. 2c; Supplementary Table 3), from 310 to 1 362 ppm, a very similar range to that recently found 248 by Luo et al. (2020) in a very large set of oil palm individuals characterised to identify the molecular 249 determinants of CPO vitamin E content. Taken together, the results of our study and those of Luo et 250 al. (2020) make it possible to propose that the range for the tocochromanol content of CPO is closer 251 to 200-1 400 ppm than to 500-800 ppm (Sundram, Sambanthamurthi, & Tan, 2003) or 600-1 000 252 ppm (Hoe, Chan, Ramanan, & Ooi, 2020; Goh, Choo, & Ong, 1985). We also observed very high 253 variability of the proportion of tocotrienols in the tocochromanols accumulated in the oil palm 254 mesocarp (Fig. 2d). The CPO of four genotypes contained a substantial proportion (35-45%) of 255 tocopherols, presenting a different picture of CPO tocochromanol composition from that previously 256 reported, in which tocotrienols largely prevailed (> 80%; Monde et al., 2009; Peh, Tan, Liao, & Wong, 257 2016; Irias-Mata et al., 2017). Given the pharmacological differences between tocopherols and 258 tocotrienols (Peh, Tan, Liao, & Wong, 2016; Trela & Szymanska, 2019; Wallert, Börmel, & Lorkowski, 259 2021), the present work provides a valuable basis for the identification of genotypes producing 260 tocopherol- or tocotrienol-rich CPO. Moreover, we found no correlation between the tocochromanol 261 content of CPO and the percentage of tocotrienols (Fig. 3), suggesting that these two traits are 262 controlled by distinct genetic determinants.

263 Using the relative percentages of the different tocochromanols, it is possible to estimate the 264 activities of the enzyme VTE3 (2-methyl-6-phytyl-1,4-hydroquinone methyltransferase), which 265 regulates competition between the biosynthetic branch of y- and α -tocochromanols and that of δ -266 and β -tocochromanols, and the enzyme VTE4 (tocopherol O-methyltransferase), which converts y-267 and δ -tocochromanols into α - and β - tocochromanols, respectively (Hunter & Cahoon, 2007; Munoz 268 & Munné-Bosch, 2019; Supplementary Fig. 1). VTE3 activity was always very high (> 88%) in the panel 269 of genotypes studied here (in other words, α - and γ -tocochromanols were always the major forms of 270 vitamin E; Supplementary Fig. 2). By contrast, VTE4 activity varied considerably among genotypes, 271 ranging from 29% in genotype AbaC4 to 74% in genotype Pobe4 (Supplementary Table 3). As the

272 bioavailability and antioxidant activity of α - and γ -tocochromanols differ significantly (Peh, Tan, Liao, 273 & Wong, 2016), the data presented here may be very important in nutritional considerations of CPO 274 intake. However, a highly significant negative correlation was found between the percentage of 275 tocotrienols and VTE4 activity (Fig. 3), suggesting that in *E. guineensis*, the enzyme VTE4 converts γ -276 tocopherol into α -tocopherol more efficiently than γ -tocotrienol into α -tocotrienol.

277 Because the tocochromanol and carotene biosynthesis pathways share the same isoprenoid 278 building blocks, they may be in competition (Asensi-Fabado & Munné-Bosch, 2010). No significant 279 correlation was found between total tocochromanol content and total carotene content in our panel 280 of 44 genotypes (Fig. 3), suggesting that there is no competition for geranylgeranyl-PP, the branch-281 point between the two pathways (Supplementary Fig. 1). This result also suggests that the activation 282 of the upstream pathway, which produces geranylgeranyl-PP in genotypes that accumulate large 283 amounts of either carotenes or tocochromanols, does not necessarily influence the two downstream 284 pathways. However, this analysis also revealed that several genotypes had high concentrations of 285 both carotenes and tocochromanols, suggesting it is possible to select varieties with high provitamin 286 A and vitamin E CPO.

287 Palmitic (16:0) and oleic (18:1 n-9) acids were the major FA in all genotypes (Fig. 2f and 2g; 288 Supplementary Table 4), followed by stearic (18:0) and linoleic (18:2) acids (Fig. 2h and 2i). Other 289 minor FA (<1%), including vaccenic (18:1 n-7), linolenic (18:3) and arachidic (20:0) acids, were also 290 detected (Supplementary Table 4). Considerable intraspecific variation was found in the levels of 291 16:0 and 18:1 (Fig. 2f and 2g), higher than that reported previously (Gascon & Wuidart, 1975; 292 Montoya et al., 2014). Indeed, although a few studies previously pointed to a low 16:0 level in some 293 oil palm varieties, e.g. 31% in Monde et al. (2009), we did not expect to identify genotypes with 25-294 26% of 16:0. Percentages of 16:0 and 18:1 were highly (R=0.92) negatively correlated 295 (Supplementary Fig. 3), as previously reported (Montoya et al., 2015). Moreover, the percentage of 296 18:0 was also negatively correlated with that of 16:0 (Supplementary Fig. 3). This finding may have 297 consequences for the melting temperature of low 16:0 CPO, since 18:0 also increases the melting 298 point of triacylglycerols. In palm oil, 18:0 is mostly incorporated in PSO triacylglycerols (P=16:0, 299 S=18:0, O=18:1; Chen et al., 2007) and PSO has a melting point as high as that of PPO (Berry, 2009). 300 The 18:2 level was not correlated with that of any of the other FA, in agreement with the distinct 301 pathways involved in the biosynthesis of 16:0, 18:0 and 18:1 in the plastid on one hand, and of 18:2 302 in the endoplasmic reticulum on the other (Dussert et al., 2013).

303 Because tocochromanols and carotenoids are strong lipid antioxidants, they are believed to be 304 accumulated by plants in seeds and fruits to protect polyunsaturated FA against peroxidation. For 305 instance, a significant relationship was found between the α -tocopherol content and the percentage of 18:2 in 14 oil crops (Kamal-Eldin & Andersson, 1997). Within our representative sample of oil palm
 diversity, neither total carotene content nor tocochromanol content was significantly correlated with
 the percentage of 18:2 (Fig. 3).

309

310 3.2. Differences in crude palm oil composition among oil palm origins

311 Significant effects of the country of origin (P=0.0067) and of the population of origin (P=0.0092) on 312 the total carotene content of CPO were observed (Fig. 4), although within-country and within-313 population variability was high in some origins: e.g., 245-992 ppm in Aba-Calabar (Nigeria). Two 314 populations contained only genotypes with high (>500 ppm) carotene content, Yocoboué in Côte 315 d'Ivoire and Pobè in Benin (Fig. 4). However, the populations La Mé (Côte d'Ivoire), Aba-Calabar 316 (Nigeria) and Deli Dabou also contained genotypes that produced a CPO rich in carotenes and were 317 thus categorised at an intermediate level according to the Duncan test. Our results are consistent 318 with those of the few previous studies which compared a limited number of origins. CPO from some 319 Nigerian genotypes had high carotene contents (Ames, Raymond, & Ward, 1960) and the variety 320 from Côte d'Ivoire analysed by Monde et al. (2009) had a higher carotene content than the variety in 321 the Deli group. No significant differences between the different groups were observed in the β -322 carotene to α -carotene ratio (Supplementary Fig. 4).

323 Although significant (P=0.04), the effects of the country and of the population of origin on the 324 CPO tocochromanol content were lower than those found for carotene content (Fig. 4). However, the 325 two populations from Côte d'Ivoire, La Mé and Yocoboué, were again distinguishable because they 326 contained genotypes with very high tocochromanol contents (ca. 1 000 ppm). The Deli Dabou group 327 was classified in an intermediate position, with several genotypes with high tocochromanol contents 328 (> 800 ppm), in agreement with the high level found in the Deli genotype investigated by Monde et 329 al. (2009). The dispersion of the percentage of tocotrienols was high in some populations, including 330 in La Mé (Côte d'Ivoire) and Yangambi (Congo) (Fig. 5). However, the effect of the country of origin 331 was highly significant (P=0.0062) and the chance of identifying varieties with a substantial percentage 332 of tocopherols appears to be greater in Côte d'Ivoire and Benin, while the Deli group showed 333 homogeneously high CPO tocotrienol levels.

The most spectacular difference between the origins studied was in 16:0 and 18:1 contents (P=0.0000 for both country and population effects; Fig. 5 and Supplementary Fig. 5). The population La Mé from Côte d'Ivoire was clearly separated from all the others with a 16:0 content < 35% in all genotypes (Fig. 5). This finding is in agreement with previous observations by Monde et al. (2009) and Gascon and Wuidart (1975). However, it is worth noting that only one genotype from Yocoboué, the other population from Côte d'Ivoire, had a low palmitate content, highlighting the specificity ofthe CPO FA composition in the La Mé population.

341 Discriminant analysis and canonical analysis were also used to assess the accuracy of classification 342 based on carotene, tocochromanol and fatty acid composition. Highly significant classifications were 343 obtained when either the population or the country of origin was used as the grouping variable, as 344 estimated by P values (<0.0000) associated with Wilks' lambda coefficients (0.00037 and 0.00250, 345 respectively). Percentages of correct classifications were very satisfactory with both grouping 346 variables (Supplementary Fig. 6). Among the 44 palm genotypes analysed, only two were 347 misclassified in both analyses. The CF1-CF2 score plots obtained by canonical analysis of CPO 348 composition clearly separated countries or populations from one another (Supplementary Fig. 6).

349 On the basis of the carotene, tocochromanol and FA composition of CPO, the material from Côte 350 d'Ivoire was distinguished from the other oil palm origins among the germplasm we studied. Using 351 microsatellite genetic markers, Cochard et al. (2009) first showed that populations from Côte d'Ivoire 352 form a distinct genetic group. Their genetic divergence likely results from a major arid phase in the 353 Dahomey gap to the west of Benin between 20000 and 14000 BP. Our data suggest that this genetic 354 separation of West African populations also led to changes in the composition of the oil stored in the 355 fruits of E. guineensis. For this reason, it would be interesting to explore the CPO composition of 356 other group I origins, such as populations from Senegal or Guinea. Based on our recent study of fruit 357 lipid composition in 144 palm species, we hypothesise that these changes reflect genetic drift rather 358 than adaptation (Guerin et al., 2020). However, one cannot exclude a role for carotenes as pigments 359 to attract dispersers, assuming that the fauna differed on the two sides of the Dahomey gap. Genetic 360 studies also provide evidence that the Deli group (group III) derived from group II, which 361 encompasses Benin, Nigeria, Congo and Angola (Cochard et al., 2009; Bakoumé et al., 2015). The 362 composition of CPO fits this hypothesis, since no major chemical differences were identified that 363 distinguished the Deli genotypes from those from these four countries.

364

365 3.3. Identifying genotypes that satisfy CPO consumers and may help in programmes to fight vitamin
 366 deficiency

Cheyns, Bricas, and Aka (2004) reported that CPO consumers in Côte d'Ivoire prefer fruits harvested in native palm groves that produce fluid and deep coloured CPO to fruits produced by modern varieties. This traditional usage was confirmed by our data, which showed that genotypes from Côte d'Ivoire had low 16:0 and high carotene contents (Fig. 4 and 5). This prompted us to investigate whether some origins combine high levels of provitamin A and vitamin E and a low level of 16:0. We 372 sorted the 44 genotypes according to decreasing carotene contents and retained those with a 16:0 373 content of less than 35% (Table 1). As expected, eight of the nine genotypes that satisfied the 16:0 374 threshold used were from Côte d'Ivoire. Among these eight genotypes, two genotypes from La Mé 375 and one from Yocoboué had both carotene and tocochromanol contents higher than 500 ppm. 376 Therefore, oil palm populations from Côte d'Ivoire represent a valuable reservoir to identify varieties 377 that satisfy both traditional CPO demand for cooking (Cheyns, Bricas, & Aka, 2004; Rafflegeau, 378 Nanda, & Genot, 2018; Rébéna, Rafflegeau, Kanski, Nanda, & Genot, 2019) and the requirements of 379 action programmes against vitamin A deficiency (Zagré, Delpeuch, Traissac, & Delisle, 2003; Burri, 380 2012), combined with increasing vitamin E intake. For instance, the genotype Lame1 showed ca. 700 381 ppm carotenes, 1 200 ppm tocochromanols and a total saturated FA percentage lower than 40%.

382 However, none of genotypes from populations of restricted origin combined the extreme values 383 found in each of the three classes of molecules: > 1 000 ppm for both carotenes and tocochromanols 384 and < 35% for 16:0. Consequently, we further investigated 12 hybrids maintained in the CRA-PP field 385 genebank at Pobè, Benin (Supplementary Tables 2-4). The hybrids were chosen because one of their 386 two parents originated from populations that were not represented in the genebank (Fig. 1): Ekona, 387 Mayang and Widikum in Cameroon, Eala and Sibiti in Congo, and a population from Nigeria, whose 388 exact location remains uncertain (and is therefore simply called 'Nigeria'). Two of the 12 hybrids had 389 particularly high carotene contents together with a low level of palmitate (Table 1). They both 390 originated from a cross between individuals from the populations La Mé and Nigeria. Moreover, the 391 hybrid LamexNige1 had the highest carotene content (1 564 ppm) among the 56 genotypes 392 evaluated, exceptionally high vitamin E content (1 330 ppm) and low total saturated FA (38.4%), 393 since it combined low 16:0 (32.3%) and low 18:0 (6.1%) levels. Crosses between La Mé and Nigeria 394 origins thus offer the opportunity to both meet CPO consumer requirements and have very high 395 vitamin contents.

396

397 **4. Conclusions**

Given the major economic importance of oil palm, information on the natural diversity and nutritional potential reported in this study should encourage the oil palm community to continue efforts to characterise CPO composition in the other major oil palm gene banks around the world, including that maintained in Malaysia (Bakoumé et al., 2015). To date, breeding efforts in oil palm have been almost entirely focused on yield and disease resistance (Murphy, 2007). Although successful in terms of these traits, they may have led to a loss in oil fluidity and in concentrations of provitamin A carotenoids in modern varieties. Our investigations showed that crosses between selected origins can combine several key nutritional properties of CPO, not only with respect to vitamins, but also its saturated FA level, whose role in health is frequently questioned and could improve consumer acceptance in Western countries (Murphy, 2007). It is also worth noting that the selection of oil palm varieties combining low palmitate and high provitamin A and vitamin E contents would facilitate both the production of red palm olein and the extraction of CPO phytonutrients (Hoe, Chan, Ramanan, & Ooi, 2020).

411

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418

419 Author contributions

Stéphane Dussert: Conceptualization, Funding acquisition, Methodology, Supervision, Writing Original Draft. Fabienne Morcillo: Conceptualization, Funding acquisition, Methodology, Supervision,
 Writing - Review & Editing. Virginie Vaissayre: Methodology, Investigation. Julien Serret:
 Investigation. Sylvie Avallone: Methodology, Investigation. Hubert Domonhédo: Resources. Florence
 Jacob: Conceptualization.

425

426 **Declaration of interest**

- 427 All authors declare no conflicts of interest exist.
- 428

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536

537 Figure captions

Figure 1. Origin of the oil palm genotypes studied here. The natural range of *E. guineensis* corresponds to the area hatched in blue. The populations collected and the number of genotypes per population studied are given in the green boxes. The size of the two subgroups of the Deli group, whose exact African origin is unknown, is given in the grey box. Hybrids between populations of different countries are indicated in the blue boxes.

Figure 2. Distribution of CPO (a) carotene content, (b) β -carotene to α -carotene ratio, (c) tocochromanol content, (d) tocotrienol percentage (e) VTE4 activity, and (f) 16:0, (g) 18:1, (h) 18:0 and (i) 18:2 levels in the 44 genotypes chosen to represent the geographic and genetic diversity of oil palm.

Figure 3. Relationships between the different components of the carotene, tocochromanol and fatty
acid composition of CPO in the 44 genotypes representative of the geographic and genetic diversity
of oil palm.

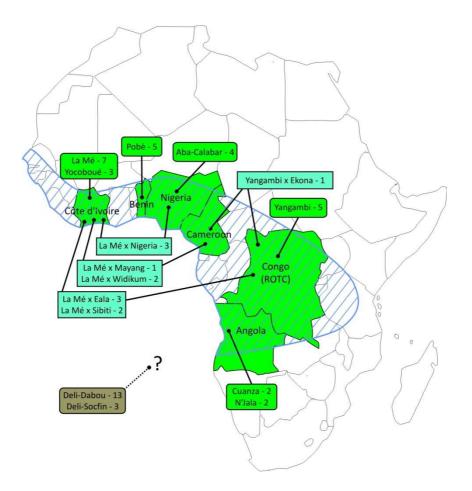
Figure 4. Effects of the country and the population of origin on CPO carotene and tocochromanol
 contents. Countries or populations followed by the same letter do not differ significantly according to
 Duncan's post hoc test.

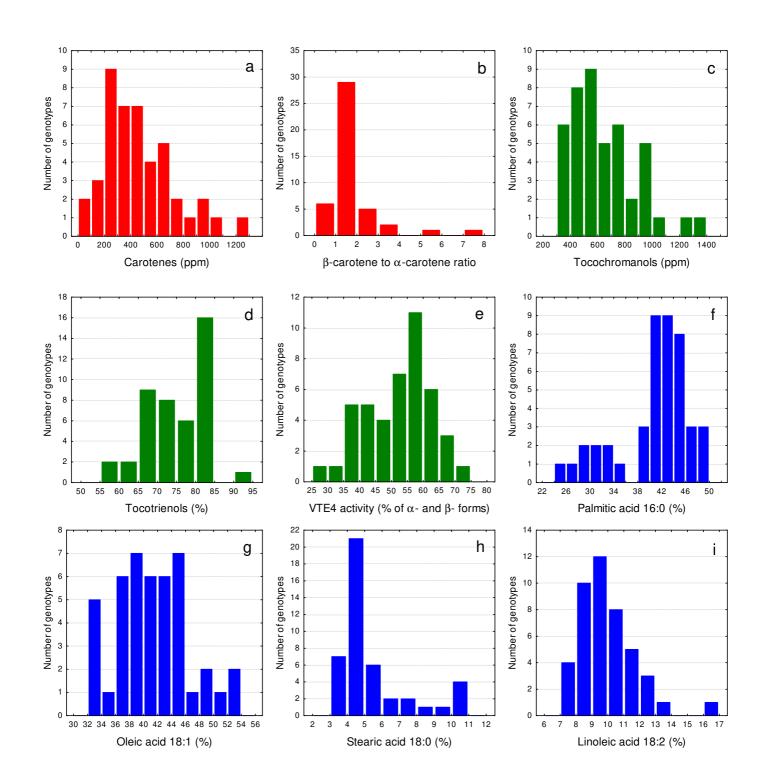
Figure 5. Effects of the country and the population of origin on CPO tocotrienol and 16:0 percentages. Groups, countries, or populations followed by the same letter do not differ significantly according to Duncan's post hoc test.

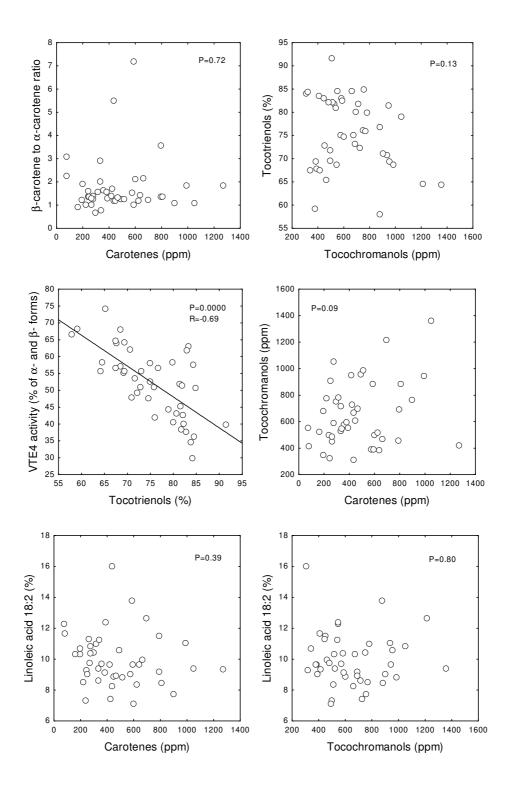
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Table 1. CPO composition of the 12 genotypes with a palmitate level below 35%. α -C and β -C: α -carotene and β -carotene. Tocols: tocochromanols. PRO: population of restricted origin.

Туре	Genotype	Carotenes	β-C to α-C	Tocols	Tocotrienols	VTE4	16:0	18:0	18:1	18:2
		(ppm)	ratio	(ppm)	(%)	activity (%)	(%)	(%)	(%)	(%)
Hybrid	LameXNige1	1564	1.5	1330	70.2	58.5	32.3	6.1	49.0	10.7
Hybrid	LameXNige2	1172	1.7	756	66.4	55.9	33.5	6.5	48.4	9.6
PRO	Lame1	698	1.2	1214	64.6	58.2	32.1	7.5	45.6	12.6
PRO	Lame7	601	2.1	495	69.5	55.6	28.8	10.4	51.9	7.1
PRO	Lame4	590	1.0	881	57.9	66.5	30.5	8.4	45.7	13.8
PRO	Үосо3	515	1.2	985	68.6	57.0	30.2	10.2	49.1	8.8
PRO	Lame2	493	1.3	956	69.3	55.2	26.6	7.6	53.5	10.6
PRO	Lame5	439	5.5	310	83.9	34.5	28.1	9.6	44.8	16.0
PRO	Lame3	338	2.9	526	81.7	45.2	25.7	10.9	52.3	9.4
PRO	Lame6	243	1.4	498	71.7	53.6	33.0	10.3	47.8	7.3
Hybrid	LameXSibi1	207	0.5	486	79.3	49.0	32.2	7.4	47.4	11.3
PRO	Cuan1	199	1.9	678	75.1	52.4	34.8	4.2	48.6	10.3







Genetic	Country	Population	Carotenes (ppm)									
group	effect P=0.0067 **	effect P=0.0092 **)	200	40	00	600	800	1000	1200		
Group I	Côte d'Ivoire ab	La Mé ^{ab}		0	0	00	O C)			···· -	
		Yocoboué ^b	-			C)	0	0		-	
Group II	Benin ª	Pobè ^b	 - 	· · · · · · · · · ·			00	0		0		
	Nigeria ^{abc}	Aba-Calabar ab	-	00	0		0		0		-	
	Congo ∘	Yangambi ª	0	0	0		0				-	
	Angola bc	Cuanza ª	-	0	0						-	
		N'Jala ª	•	С) C)						
Group III	Deli ^{bc}	Deli Dabou ^{ab}	-	00	00			0 0	C			
		Deli Socfin a	· .	0 0	6	0					-	
				1						4		

Genetic group	Country effect P=0.0400 *	Population effect P=0.0430 *	4	00	Toc 600			ols ((ppm) 1200	1400	
Group I	Côte d'Ivoire a	La Mé ^{ab}	0 0		0 0		0 0		0		····· -
		Yocoboué ^b	-			0		0		0	8-
Group II	Benin ^b	Pobè a	- (000	>						
	Nigeria ^{ab}	Aba-Calabar a	0		0	0	0				-
	Congo b	Yangambi ª	α	∞	0	0					-
	Angola ab	Cuanza ª	-		0	0					
		N'Jala ª	-		∞						
Group III	Deli ^{ab}	Deli Dabou ^{ab}	-	0	00		0	0			
		Deli Socfin a	00		∞	C		0			

Genetic	Country	Population effect	Tocotrienols (%)								
group	effect P=0.0062 **	P=0.0378 *	60	65	70	75	80	85	90		
Group I	Côte d'Ivoire a	La Mé ^{ab}	• 0	0	0 00		00				
		Yocoboué b	-	0	0	0			-		
Group II	Benin ª	Pobè ^b	-	0	00	0			-		
	Nigeria ^b	Aba-Calabar ab	-		0	0		0	0		
	Congo ab	Yangambi ^{ab}	0		0		0	∞	-		
	Angola ab	Cuanza ab	-			0			-		
		N'Jala ab	-		0		i)	0	(-		
Group III	Deli ^b	Deli Dabou ^{ab}	ංකා කර ඉති කර					DO	-		
		Deli Socfin ^a)	-		

Genetic	Country	Population	Palmitic acid 16:0 (%)								
group	effect P=0.0000 ***	effect P=0.0000 ***	25	30	35	4	10	45	50		
Group I	Côte d'Ivoire a	La Mé ª	- 00 0	$\infty \circ \infty$							
		Yocoboué ^b		0		0	0 		-		
Group II	Benin bc	Pobè ∘	-			0	0	8	0		
	Nigeria bc	Aba-Calabar º	-				0	0	<u>o</u> -		
	Congo c	Yangambi °	-				0	C	00 -		
	Angola ^b	Cuanza ^b	-		0		0		-		
		N'Jala ∘	-				0	0	-		
Group III	Deli ^{bc}	Deli Dabou ∘	-			0	00	യന	-		
		Deli Socfin ∘	-				0		-		